

EGYPTIAN ACADEMIC JOURNAL OF BIOLOGICAL SCIENCES TOXICOLOGY & PEST CONTROL



ISSN 2090-0791

WWW.EAJBS.EG.NET

Vol. 13 No. 2 (2021)

EABS BS Egypt. Acad. J. Biolog. Sci., 13(2:49-70(2021)

Egyptian Academic Journal of Biological Sciences F. Toxicology & Pest Control ISSN: 2090 - 0791 http://eajbsf.journals.ekb.eg/



# Suppressive Impact of 6-Benzyladenine, A Plant Growth Regulator, on the Adult Performance and Reproductive Potential of *Galleria mellonella* L. (Lepidoptera: Pyralidae)

Abo Elsoud, A.A.<sup>1</sup>; Ghoneim, K.<sup>1\*</sup>; Hamadah, Kh.<sup>1</sup>and El-Hela, A.<sup>2</sup>

1- Departmentof Zoology and Entomology, Faculty of Science, Al-Azhar University, Cairo, Egypt
2- Faculty of Pharmacy, Al-Azhar University, Cairo, Egypt
E-mail\* :<u>karemghoneim@gmail.com</u>

#### ARTICLEINFO

Article History Received: 25/5/2021 Accepted: 15/7/2021

*Keywords*: Cytokinin, longevity, oviposition, postoviposition, preoviposition, incubation, fecundity, fertility, sterility.

### ABSTRACT

The greater wax moth Galleria mellonella is responsible for serious economic losses to bee keepers in developing countries. Larvae feed on the wax comb in weak colonies or during the storage of wax combs in winter. The objective of the current study was to assess the effect of the Cytokinin plant growth regulator, Benzyladenine (6-BA), on the most important parameters of adult performance and reproductive potential of G. mellonella. The 3<sup>rd</sup> instar larvae were force-fed on an artificial diet supplemented with six concentrations, viz., 100, 10.0, 1.0, 0.1, 0.01 & 0.001ppm of 6-BA. No adult moths could metamorphose at the higher three concentrations because of larval and pupal deaths. The most important results could be summarized as follows. The adult emergence was blocked by 6-BA, in a dose-dependent course. Also, the tested compound displayed strong adulticidal activity, since adult mortality increased in a dose-dependent course. The total adult longevity of moths was significantly prolonged.All longevity compartments were remarkably prolonged. On the other hand, 6-BA failed to affect adult morphogenesis. The oviposition efficiency was deleteriously prohibited. Also, 6-BA exhibited a tremendous inhibitory effect on the female fecundity and reducing effect onfertility. The embryonic development in eggs laid by treated adult females was severely retarded, since the incubation period of these eggs was remarkably prolonged.

# **INTRODUCTION**

The greater wax moth *Galleria mellonella* L. (Lepidoptera: Pyralidae) is widely distributed throughout the world. It has been recorded in more than 77 countries (Kwadha *et al.*, 2017; Roh *et al.*, 2020). It is the major destructive pest for the apiculture industry because its larvae feed on the honeycomb, honey and wax found in bee hives, especially in weak colonies or during the storage of wax combs in winter (Büyükgüzel and Kalender, 2009), in addition to their tunneling habit through the combs (Chandel *et al.*, 2003).

Different physical and chemical methods have been used to control *G. mellonella* (Büyükgüzel, 2009). These methods and materials are very expensive and may have toxic effects on the bees. Furthermore, these methods may contaminate the bee products (Rortais *et al.*, 2017). In different parts of the world, the apiculture industry has traditionally relied on synthetic insecticides for controlling carious insect pests (Rehman *et al.*, 2009; Ilyas *et* 

*al.*, 2017). These insecticides have dangerous impacts on non-target organisms and on the environment in general. They gradually accumulate in food materials and ultimately cause severe diseases in human consumers, like cancer, kidney failure and genetic disorders (Owain *et al.*, 2008; Ambethger, 2009; Łozowicka *et al.*, 2012; Mahdi and Mohammed, 2017). Also, one of the serious problems that emerged by the synthetic insecticide applications is the development of insect resistance toward the used insecticides (Yarahmadi *et al.*, 2009; Sparks and Nauen, 2015; Maazoun *et al.*, 2017). *G. mellonella* has developed high resistance to many insecticides (Said *et al.*, 2019).

Therefore, new environmentally safer alternative compounds should be searched (Ansari *et al.*, 2012; Glare *et al.*, 2016; Liao *et al.*, 2017; Kunbhar *et al.*, 2018). The use of natural products is a promising alternative for controlling *G. mellonella* because they generally have a low environmental contamination risk and there are few if any, harmful residues in the bee products (El-Wakeil, 2013; Farghaly *et al.*, 2017). Recently, the plant growth regulators (PGRs) have received increasing attention because they are environmentally friendly compounds of the plant origin (Abdellaoui *et al.*, 2015; Er and Keskin, 2016). Many authors (Kaur and Rup, 2002; Mendonça *et al.*, 2006; Gupta *et al.*, 2009; Tsagkarakis *et al.*, 2012; Prado and Frank, 2013; Kaur *et al.*, 2016) reported that PGRs may have drastic effects on the survival, development and reproductive potential, as well as disturbance of other physiological processes and induction of the oxidative stress. Various authors (Altuntaş *et al.*, 2012; Uçkan *et al.*, 2014; Altuntaş, 2015), also, suggested that PGRs, such as gibberellic acid, ethephon and indole-3-acetic acid, could be used instead of synthetic insecticides to control some lepidopterous pests.

PGRs have been classified into different categories. Hopkins and Hüner (2004) classified the PGRs into six classes: Gibberellins (GAs), Auxins (Auxs), Ethylene (ET), Cytokinins (CTKs), Abscisic acid (ABA) and Brassinosteroids (BRs). Stamm *et al.* (2011) classified the PGRs into main nine classes: Auxs, GAs, CTKs, ET, ABA, BRs, salicylic acid (SA), Jasmonates (JAs) and Strigolactones (SLs). It is well known hitherto that CTKs have been produced in almost all higher plants as well as many mosses and prokaryotes (Salisbury and Ross, 1992). In addition, insects may produce CTKs, either directly or indirectly owing to their association with endosymbiotic bacteria (Giron and Glevarec, 2014; Zhang *et al.*, 2016). In insects, also, CTKs have been detected in saliva, labial glands, or/and accessory glands suggesting their ability to produce and deliver these compounds into plants as PGRs (Giron *et al.*, 2013; Body et al., 2013; Bartlett and Connor, 2014). However, CTK has been reported to affect the morphology, development and behavior of some insects (Rup *et al.*, 1998).

In this context, the 6-Benzyladenine (6-BA) (or 6-Benzylaminopurine, BAP) is one of the most important synthetic CTKs. It is a plant growth stimulator with the chemical name: 4-hydroxyphenethyl alcohol. In plants, 6-BA can significantly increase CTK levels, of which levels are dramatically diminished under stress. Applying 6-BA was conducive to minimize adverse effects of environmental stress, such as drought, salt, low-temperature, and waterlogging stress (Majid *et al.*, 2011; Nurunnaher *et al.*, 2014; Müller and Theron, 2018). For uses of 6-BA in agriculture, see Mangena (2020), Hu *et al.* (2020) and Bubán (2020). The objective of the current study was to assess the effect of 6-BA on the most important parameters of adult performance and reproductive potential of *G. mellonella*.

#### **MATERIALS AND METHODS**

#### The Insect:

A culture of asusceptible strain of the greater wax worm *Galleria mellonella* L. (Lepidoptera: Pyralidae) was established in the Department of Zoology, Faculty of Science, Al-Azhar University, Cairo, Egypt, and maintained for several successive generations under controlled conditions  $(27\pm2^{\circ}C, 65\pm5\%$  R.H., photoperiod 14 h L and 10 h D). This culture was originally initiated by a sample of larvae kindly obtained from Desert Research Center, Cairo, Egypt. Larvae were transferred into glass containers, tightly covered with the muslin cloth. Different techniques for preparing the artificial diet had been described by some authors (Metwally *et al.*, 2012; Nitin et al., 2012). In the present culture of *G. mellonella*, an artificial diet was formulated depending on the method of Bhatnagar and Bareth (2004). The diet contained maize flour (400 g), wheat flour, wheat bran and milk powder, 200 g of each. Also, it was provided with glycerol (400g), bee honey (400g), yeast (100g). However, improved manipulation of different developmental stages had been done according to Ghoneim *et al.* (2019 a).

### 2. Plant Growth Regulator and Larval Treatment:

The compound 6-Benzyladenine (6-BA, or 6-Benzylaminopurine, BAP) is a plant growth stimulator (Cytokinin class) with the chemical name: 4-hydroxyphenethyl alcohol and molecular formula:  $C_{12}H_{11}N_5$ . It was purchased from Milipore Sigma, Burlington, MA 01803, USA Merk Ltd., Cairo, Egypt. A series of six concentration levels of 6-BA was prepared by diluting the compound with distilled water in volumetric flasks, as follows: 100.0, 10.0, 1.0, 0.1, 0.01 and 0.001 ppm.

Ten grams of the previously described artificial diet were supplemented with 2 ml of each concentration of 6-BA before introduction to the newly moulted  $3^{rd}$  instar larvae, as a food. These larvae were allowed to feed continuously on the treated diet throughout the larval stage. Control larvae were provided with distilled water-treated diet. Ten replicates of treated and control larvae (one larva/replicate) were kept separately in suitable glass vials under controlled laboratory conditions ( $27\pm2^{\circ}$ C,  $65\pm5\%$  R.H., photoperiod 14 h L and 10 h D). After force-feeding of  $3^{rd}$  instar larvae of diet supplemented with the higher three concentrations (100.0, 10.0 & 1.0ppm), larvae and pupae died. Therefore, all parameters of adult performance and reproductive potential were determined after feeding of similar larvae on diet mixed with the lower three concentrations of 6-BA (0.1, 0.01 & 0.001ppm).

# Criteria of Study:

#### **1. Adult Performance**:

Adult Emergence: The number of the successfully emerged adults was expressed in %, according to Jimenez-Peydro *et al.* (1995), as follows:

[No. of completely emerged adults / No. of pupae] ×100

Adult Mortality: It was calculated in % of the adult moth deaths.

Adult Morphogenesis: The impaired morphogenesis program of adult transformation was expressed in % of adult deformities.

Adult Longevity: The most important compartments of the longevity of adult females were measured in days ( $\pm$ SD): pre-oviposition period, oviposition period (reproductive lifetime) and post-oviposition period.

# 2. Reproductive Potential:

After feeding of 3<sup>rd</sup> instar larvae on an artificial diet (treated with 6-BA or control), the emerged adult moths were kept separately in glass jars (3 L) provided with white paper scraps, as oviposition sites. Each adult female was coupled with normal adult males (1:2) of the same age, obtained from the main culture. After mating, female moths were allowed to

lay eggs. The eggs, singly or in patches, were collected daily, and carefully transferred into Petri dishes to be counted.

**The Oviposition Efficiency**: The oviposition efficiency was denoted by the **oviposition rate** which was calculated as follows:

Number of laid eggs per  $\mathcal{Q}$ / reproductive lifetime (in days)

**The Reproductive Capacity:** The most important parameters of reproductive capacity were fecundity and fertility. **Fecundity:** the laid eggs were counted for calculating the number of eggs per female. **Fertility:** the hatchability was usually expressed in the hatching percentage of the laid eggs. **The sterility index** was calculated according to Toppozada*et al.*(1966) as follows:

*Sterility Index* = 100 – [(*a b* / *A B*) × 100]

Where: a: mean number of eggs laid per female in the treatment. b: percentage of hatching in the treatment. A: mean number of eggs laid per female in the controls. B: percentage of hatching in the controls.

### **3. Incubation Period:**

The laid eggs were kept in Petri dishes under the previously mentioned laboratory conditions. Just after the oviposition, eggs were observed until hatching to determine the incubation period (in days±SD).

# 4. Data Analysis:

Data obtained were statistically analyzed by the student's *t*-distribution, and refined by Bessel correction (Moroney,1956) for the test significance of the difference between means using GraphPadInStat<sup>©</sup> v. 3.01 (1998).

### RESULTS

# I. Effect of 6-Benzyladenineon the Adult Performance of G. mellonella:

After force-feeding of the  $3^{rd}$  instar larvae of *G. mellonella* on diet mixed with six concentrations of 6-Benzyladenine (6-BA) (100, 10.0, 1.0, 0.1, 0.01 & 0.001ppm), no adults could metamorphose at the higher three concentrations. Therefore, the adult performance parameters could be determined after larval feeding only on three sublethal concentrations. Data of these parameters were summarized in Table (1).

# 1.1. Effect of 6-BA on the Adult Emergence:

In the light of data assorted in the previously mentioned table, the adult emergence was blocked, in a dose-dependent course (20, 60 & 80% emergence of treated adults, at 0.1, 0.01 & 0.001ppm, respectively, compared to 100% emergence of control adults).

# 1.2. Effect of 6-BA on Adult Survival:

Depending on the same table, 6-BA displayed strong adulticidal activity, since adult mortality increased by the increasing concentration (80, 40 & 20% mortality of treated adults, at 0.1, 0.01 & 0.001ppm, respectively, compared to 0% mortality of control adults).

# 1.3. Effect of 6-BA on Adult Morphogenesis:

No deformed adults could be observed, i.e., the 6-BA failed to affect the adult morphogenesis.

### **1.4. Effect of 6-BA on Adult Longevity:**

As obviously shown in Table (1), the total adult longevity was significantly prolonged, in a dose-dependent course  $(17.2\pm0.28, 16.4\pm0.30 \& 14.2\pm0.34 \text{ days of treated}$  adults, at 0.1, 0.01 & 0.001ppm, respectively, *vs.* 13.2±0.30 days of control adults). One of the major time compartments of adult longevity is the pre-oviposition period (may be theovarian maturation period). This period was generally prolonged. Moreover, it was pronouncedly prolonged at the highest concentration (12.1±0.42, 3.0±0.14 & 3.4±0.42 days of treated adults, at 0.1, 0.01 & 0.001ppm, respectively, *vs.* 3.2±0.35 days of control adults).

Another major time compartment of adult longevity is the oviposition period (reproductive lifetime). Also, this period was considerably prolonged ( $7.0\pm0.42$ ,  $6.1\pm0.14$  &  $5.7\pm0.70$  days of treated adults, at 0.1, 0.01 & 0.001ppm, respectively, *vs.*  $5.5\pm0.56$  days of control adults). In addition, the last time compartment is the post-oviposition period. It was remarkably prolonged ( $7.2\pm0.14$ ,  $6.5\pm0.14$  &  $6.1\pm0.70$  days of treated adults, at 0.1, 0.01 & 0.001ppm, respectively, *vs.*  $4.6\pm0.28$  days of control adults, Table 1).

**Table1**: Effect of 6-Benzyladenine on the adult performance parameters of *G. mellonella* after force-feeding of 3<sup>rd</sup> instar larvae on treated artificial diet.

			Adult longevity (mean days±SD)				
Parameter Conc. (ppm)	Adult emergence (%)	Adult mortality (%)	Pre-oviposition period (Ovarian maturation period)	Oviposition period (Reproducti ve life-time)	Post- oviposition period	Total Longevity	
0.1	20	80	12.1±0.42 c	7.0±0.42 c	7.2±0.14 d	17.2±0.28 d	
0.01	60	40	3.0±0.14 a	6.1±0.14 a	6.5±0.14 d	16.4±0.30 d	
0.001	80	20	3.4±0.42 a	5.7±0.70 a	6.1±0.70 c	14.2±0.34 c	
Control	100	00	3.2±0.35	5.5±0.56	4.6±0.28	13.2±0.30	

Conc.: concentration. Mean $\pm$ SD followed with (a): insignificantly different (P>0.05). (c): highly significantly different (P<0.01),(d): very highly significantly different (P<0.001).

# 2. Effect of 6-BA on the Reproductive Potential:

After force-feeding of  $3^{rd}$  instar larvae of *G. mellonella* on an artificial diet supplemented with six concentrations of 6-BA (100, 10.0, 1.0, 0.1, 0.01 & 0.001ppm), successfully emerged and reproduced adult female moths were observed only at the three lower concentrations. Data of the most important criteria of the reproductive potential were summarized in **Table (2)**.

### 2.1. Effect of 6-BA on Oviposition Efficiency:

According to data of this table, the oviposition efficiency was deleteriously prohibited by 6-BA, since the oviposition rate was drastically regressed, in a dose-dependent course ( $26.6\pm0.28$ ,  $33.4\pm0.56$  &  $42.0\pm1.41$  of treated adult females, at 0.1, 0.01 & 0.001ppm, respectively, *vs.*,  $45.8\pm0.56$  of control adult females).

### 2.2. Effect of 6-BA on Reproductive Capacity:

Data of the same table revealed a tremendous inhibitory effect of 6-BA on the female fecundity (mean number of eggs/ $\bigcirc$ ), in a dose-dependent manner (80.8±2.82, 121.3±1.41 & 189.5±1.41 eggs /treated $\bigcirc$ , at 0.1, 0.01 & 0.001ppm, respectively, compared to 229.9±2.82 eggs /control $\bigcirc$ , Table 2).

Another informative parameter of the reproductive capacity is fertility (hatching% of laid eggs or egg viability) which was significantly reduced,  $(63.9\pm0.14, 69.5\pm0.70 \& 70.1\pm0.14$  hatching eggs laid by treated  $\bigcirc \bigcirc$ , at 0.1, 0.01 & 0.001ppm, respectively, compared to 70.3\pm0.42 hatching eggs laid by control  $\bigcirc \bigcirc$ ). It may be important to estimate the sterilityindex which increased parallel to the increasing concentration of 6-BA (69.69, 48.21 & 18.49, at 0.1, 0.01 & 0.001ppm, respectively, Table 2).

### 2.3. Effect of 6-BA on Embryonic Development:

In insects, the incubation period of eggs can be used as an informative indicator of the embryonic developmental rate, i.e., a longer period usually denotes as lower rate of embryogenesis and *vice versa*. On the basis of data assorted in Table (2), the embryonic development in eggs laid by treated adult females was severely retarded with a noticeably slower rate, since the incubation period of these eggs was significantly prolonged ( $9.3\pm0.42$ ,

 $9.0\pm0.70$  &  $8.9\pm0.14$  days of eggs laid by treated adult females, at 0.1, 0.01 & 0.001ppm, respectively, compared to  $7.4\pm0.28$  days of eggs laid by control adult females).

Table 2:	Effects of	6-Benzyladenine	on the	oviposition	and	reproductive	capacity	of <i>G</i> .
ľ	nellonella,	after force-feedi	ng of 3 <sup>1</sup>	<sup>d</sup> instar larv	ae on	treated artifi	cial diet.	

	Ovingsition	Re	Incubation			
Conc. (ppm)	rate (mean±SD)	Fecundity (mean no. of eggs ±SD)	Fertility (mean %±SD)	Sterility index (%)	period (Mean days ±SD)	
0.1	26.6±0.28 d	80.8±2.82 d	63.9±0.14 d	69.69	9.3±0.42 d	
0.01	33.4±0.56 d	121.3±1.41 d	69.5±0.70 b	48.21	9.0±0.70 c	
0.001	42.0±1.41 c	189.5±1.41 d	70.1±0.14 a	18.49	8.9±0.14 d	
Control	45.8±0.56	229.9±2.82	70.3±0.42		7.4±0.28	

Conc., a, c, d: see footnote of Table (1). (b): significantly different (P<0.05).

### DISCUSSION

# **I.** Disruption of Adult Performance of *G. mellonella* by 6-Benzyladenine: **I.1.** Blocked Adult Emergence:

It is important to emphasize that the adult emergence in insects is a crucial physiological process and regulated by some hormones, particularly the eclosion hormone. Disturbance of this hormone partially or completely arrests the adults to emerge (Josephrajkumar *et al.*, 1999; Ghoneim *et al.*, 2019 b).

In the present study, force-feeding of  $3^{rd}$  instar larvae of G. mellonella on diet supplemented with three sublethal concentrations of 6-Benzyladenine (6-BA) led to the blockage of adult emergence, in a dose-dependent course. The present result was in agreement with many reported results of blocked adult emergence after larval treatment of various insects with some plant growth regulators (PGRs), such as the melon fruit fly Bactrocera cucurbitae after feeding of larvae on artificial diets containing different concentrations of gibberellic acid (GA<sub>3</sub>)(Kaur and Rup, 1999) or Coumarin (Cn), kinetin, GA<sub>3</sub> and indole-3-acetic acid (IAA) (Kaur and Rup, 2003); the tobacco cutworm Spodoptera *litura* by rearing the newly hatched larvae on artificial diets fortified with miraculan (Singh and Bhattacharya, 2001; Bhatnagar et al., 2012) or GA<sub>3</sub> (Shiwani and Karnatak, 2012) and the mustard aphid Lipaphis erysimi after treatment of nymphs with GA3, Daminozide(Alar-B9), indole-3-butyric acid (IBA) or Chlorogenic acid (Rup et al., 2002). On the contrary, our result disagreed with very few reported results of enhanced adult emergence of some insects after larval treatments with some PGRs, such as the Egyptian cotton leafworm Spodoptera littoralis after feeding of larvae on diet supplemented with GA3 (Salama and El-Sharaby, 1972) and G. mellonella after injection of Abscisic acid (ABA) into the haemocoel (Er and Keskin, 2016).

For interpretation of the blocked adult emergence of *G. mellonella* after forcefeeding of  $3^{rd}$  instar larvae on diet mixed with 6-BA, in the currentstudy, this compound might interfere with the eclosion hormone release and/or inhibition of the neurosecretion (prothoracicotropic hormone) (Al-Sharook *et al.*, 1991; Josephrajkumar *et al.*, 1999). The 6-BA might exhibit a disturbing effect on the normal metabolism of insect hormones during the development of the immatures leading to failure of adult emergence (Trigo *et al.*, 1988). On the molecular basis, 6-BA might cause misexpression of certain genes, particularly the brood complex (*br-C*) transcription factor gene, leading to symptoms of impaired metamorphosis, like blocking of adult emergence (Wilson, 2004; Nandi and Chakravarty, 2011).

#### I.2. Adulticidal Activity of 6-BA:

After force-feeding of 3<sup>rd</sup> instar larvae of *G. mellonella* on diet supplemented with 6-BA, in the current investigation, the compound displayed strong adulticidal activity, since adult mortality increased by the increasing concentration. In the available literature, no information was found on the chronic toxicity of PGRs against adults of insects after treatment of larvae. Apart from PGRs, different plant-derived compounds were reported to exhibit adulticidal activities against some insects, such as Thymoquinone against the maize weevil *Sitophilus zeamais* (Herrera *et al.*, 2015); *trans*-cinnamaldehyde, (–)-menthone and eugenol against the rice weevil *Sitophilus oryzae* (Saad *et al.*, 2018); Nerolidol (Hamadah *et al.*, 2020) and Farnesol (Hamadah *et al.*, 2021) against *S. littoralis*.

To interpret the chronic toxic effect of 6-BA on adult moths of *G. mellonella*, in the present study, this PGR might be retained and distributed in the body, as a result of direct and rapid transport from the gut of treated larvae into other tissues, through haemolymph, to the adults and by lower detoxification capacity of adults against the tested PGR (Osman *et al.*, 1984; Smagghe and Degheele, 1992). Because the adult life in insects depends on healthy immature stages, the digestive disorders may be the cause of untimely adult mortality, as recorded for 6-BA against *G. mellonella* adults in the current study (Soltani, 1984). Also, an extended toxic effect of 6-BA might be due to the disturbance of enzymatic pattern and/or hormonal hierarchy in adults of *G. mellonella*(Kartal*et al.*, 2003). In addition, the adult mortality of *G. mellonella* might be explicated by a latent inhibitory effect of 6-BA on the feeding leading to continuous starvation and subsequently death (Ghoneim et al., 2000) or adverse effect on the homeostasis leading to increased loss of the body water and subsequently death (Amer *et al.*, 2004).

### I.3. Disturbed Adult Longevity by 6-BA:

### I.3.1. Total Adult Longevity:

There are many reported results of shortened total adult longevity of different insects after treatment with various PGRs. For example, thetotal adult longevity of *G. mellonella* moths was shortened after feeding of larvae on diet treated with GA<sub>3</sub> (Uckan *etal.*, 2011) or mebendazole (Calık *et al.*, 2016) or by injection of ABA into the haemocoel (Er and Keskin, 2016). Also, feeding of *S. littoralis* larvae on a diet supplemented with GA<sub>3</sub> resulted in shortened total adult longevity (Salama and El- Sharaby, 1972). Exposure of the migratory grasshopper *Melanopliss anguinipes* nymphs to Ethylene led toshortened female adult longevity (Chrominski *et al.*, 1982). The total adult longevity of *B. cucurbitae* was shortened by GA<sub>3</sub> treatments (Kaur and Rup, 2002). The total adult longevity of wolfberry aphid (Gong *et al.*, 2010) or brown planthopper *Nilapar vatalugens* (Senthil-Nathan *et al.*, 2009) was shortened after treatment with Jasmonic acid (JA). In a recent study by Nagaratna *et al.* (2021), feeding of larvae of the fall armyworm *Spodoptera frugiperda* on maize treated with silicic acid (FSA), GA<sub>3</sub> and JA, separately or in combination, resulted in thes hortening of total adult male longevity by all treatments but total adult female longevity was unaffected.

The present results disagreed with the previously reported results since forcefeeding of  $3^{rd}$  instar larvae of *G. mellonella* on diet supplemented with 6-BA resulted in significantly prolonged total adult longevity, in a dose-dependent course. However, the current literature contains no information about the prolongation of total adult longevity of insects after treatment with PGRs except a recent study by Çelik and Sak (2021) who investigated the effects of kinetin against *A. grisella* and recoded significantly prolonged adult longevity of males, at 5 mg/Lkinetin. Interpretation of the shortened total adult longevityof different insects after larval treatments with various PGRs is available (Abdel-Aal, 1996; Broughton *et al.*, 2005; Carbone *et al.*, 2006; Kefford *et al.*, 2008; Chamseddin *et al.*, 2012; Er *et al.*, 2017) while the exact interpretation of prolongation of total adult longevityof *G. mellonella* after larval feeding on diet mixed with 6-BA, in the present study, is unfortunately available right now. On the other hand, insects often show degenerative changes in some tissues and organs, after the attainment of sexual maturity, which can be called 'senility' or 'aging'. The affected adult longevity can be considered as an informative indicator for adult aging, i.e., prolongation of longevity may denote a delay of aging but shortened longevity may denote an acceleration of aging, although the death is usually the density of all creatures (for review see Ghoneim and Bakr, 2018). Depending on our results, 6-BA seemed to delay the aging of *G. mellonella*, i.e., it exhibited antioxidant activity against this insect. However, the exact mode of action of 6-BA on the biochemical sites in adults is unknown until now.

#### I.3.2. Prolonged Pre-Oviposition Period:

To the best of our knowledge, no information is available on the effects of PGRs on major compartments of the adult longevity (pre-oviposition period, oviposition period and post-oviposition period) of insects, except one study conducted by Abdellaoui *et al.* (2009) on *L. migratoria* who topically applied GA<sub>3</sub> or injected it into oesophagus of the newly emerged adults and recorded a prolongation of the pre-oviposition period. Apart from PGRs, Nerolidol (Hamadah *et al.*, 2020) and Farnesol (Hamadah *et al.*, 2021) inhibited the adult female moths of *S. littoralis* to pass generally prolonged pre-oviposition period. Results of the present study were, to some extent, in agreement with those reported results, since force-feeding of  $3^{rd}$  instar larvae of *G. mellonella* on a diet mixed with 6-BA led to a general prolongation of the pre-oviposition period.

Many lepidopterous species have a relatively short adult stage, or even non-feeding adults, like *G. mellonella*. In these insects, anadult female emerges with most of her eggs ready to be fertilized within a day or very few days. The prolonged pre-oviposition period of *G. mellonella* in the present study may be informative to delaying or retarding the effect of 6-BA on the finishing steps of ovarian maturation (Ghoneim and Al-keridis, 2019). In addition, ovarian maturation in insects is known to be under endocrine control (Kaur and Rup, 2002). The delayed ovarian maturation in *G. mellonella* by 6-BA, in the current work, seemed to be related to its interference with the inhibition of ecdysteroid production in the ovaries, the occurrence of which is a prerequisite for the developing ovaries (Acheuk et al., 2012). However, the exact mode of delaying effect of 6-BA on the ovarian maturation of *G. mellonella*, in the current investigation, is unfortunately not available right now. On the other hand, the interference of this compound with the hormonal regulation of this physiological process needs further investigation in the foreseeable future.

#### I.3.3. Prolonged Oviposition Period:

Another major time compartment of adult longevity is the oviposition period (reproductive lifetime). Depending on the currently available literature, there are no studies examining the effects of PGRs on this period other than the study of Abdellaoui *et al.* (2009) who topically applied or injected different concentrations of GA<sub>3</sub> into the oesophagus of the newly emerged adult females on *L. migratoria* and recorded a reduction of the oviposition period. Aside from PGRs and *G. mellonella*, treatment of *S. littoralis* larvae with Nerolidol (Hamadah *et al.*, 2020) or Farnesol (Hamadah *et al.*, 2021) resulted in a remarkably shortened oviposition period of the successfully mated female moths.

Results of the present study clearly disagreed with those reported results, since forcefeeding of  $3^{rd}$  instar larvae of *G. mellonella* on a diet mixed with 6-BA led to considerable prolongation of the oviposition period of adult females. The shortened oviposition period can be understood as a result of enforcing the effect of the tested compound on adult females to quickly lay eggs during a very short interval to avoid this toxic xenobiotic factor (Tanani and Ghoneim, 2018) while the prolongation of this period after treatment of *G. mellonella* larvae with 6-BA, in the present study, could not be explicated right now!!

#### **I.3.4. Prolonged Post-Oviposition Period:**

The last time compartment of the adult longevity in insects is the post-oviposition period. From the current literature, no reliable information has been obtained regarding the effects of PGRs, or other plant-derived compounds, on this time interval of adult longevity in insects. Only Ghoneim *et al.* (2019 b) recorded a considerably shortened post-oviposition period of the successfully mated adult females of *G. mellonella* after treatment of  $3^{rd}$  instar larvae with the honey bee venom, Apitoxin. Our result is contradictory to this reported result since the post-oviposition period of *G. mellonella* was considerably prolonged after larval treatment with 6-BA. Unfortunately, there is no acceptable interpretation for this prolongation right now!!

#### I.4. Deranged Adult Morphogenesis by 6-BA:

In the current study, 6-BA failed to affect the adult morphogenesis, since no adult deformities of *G. mellonella* had been observed after force-feeding of 3<sup>rd</sup> instar larvae on a diet mixed with different concentrations of this PGR. In contrast to our result, very few studies examined the effects of PGRs on morphogenesis of some insects. Yeşilada *et al.* (1996) observed adult abnormalities of the vinegar fly *Drosophila melanogaster* after treatment of different developmental stages with kinetin and ABA. Kaur and Rup (2003) observed some morphological deformities in adult flies of *B. cucurbitae* after treatment of larvae with Cn, GA<sub>3</sub>, kinetin, or IAA. Recently, Çelik and Sak (2021) investigated the effects of kinetin against *A. grisella* and observed moths with curved wings, shortened wings.

Apart from PGRs, different plant-derived compounds had been reported to adversely affect the morphogenesis of some insects. For example, treatment of the confused flour beetle *Tribolium confusum* larvae with Andrographolide resulted in the production of some deformed adults (Lingampally *et al.*, 2013). Feeding of  $2^{nd}$  instar larvae of *S. litura* on fresh food treated with Allyl isothiocyanate resulted in the metamorphosis of some deformed adults (Bhushan *et al.*, 2016). Larval treatment of *S. litura* and *Spodoptera exigua* withPogostone resulted in some deformities of the adult moths (Huang *et al.*, 2014). Topical application of Farnesol onto 5<sup>th</sup> instar nymphs of the red cotton stainer *Dysdercus koenigii* led to the metamorphosis of adults with malformed wings (Kumar and Gupta, 2017). Sosa *et al.* (2019) observed serious wing malformations in adults of *S. littoralis*  $2^{nd}$  instar larvae on plant leaves previously dipped in Nano-chitosansolution led to the production of deformed adult moths (Marouf, 2020). Different malformations of *S. littoralis* adults had been observed after larval treatments with the sesquiterpene compounds, Nerolidol (Hamadah *et al.*, 2020) and Farnesol (Hamadah *et al.*, 2021).

#### II. Impaired Reproductive Potential of G. mellonella by 6-BA:

Great research attention has been focused on the drastic activities of PGRs against the reproduction of insect pests. For example, a reduction in the reproductive rate of some species of aphids was reported after feeding on some plants treated with chlormequat chloride (Chch) (Honeyborne, 1969). Mepiquat chloride (Mch) reduced the reproductive rate of wheat aphid *Schizaphis graminum* (Dreyer *et al.*, 1984). A similar result of impaired reproduction was obtained for the same aphid species on GA<sub>3</sub>-treated sorghum crops (Campbell *et al.*, 1984). Also, Cytokinin (CTK) has been reported to affect the reproduction of mustard aphid *Lipaphis erysimi* (Rup *et al.*, 2000). In addition to aphids, GA<sub>3</sub> reduced the reproductive potential of the Mediterranean fruit fly *Ceratitis capitata* (Barbouche and Ben Hamouda, 1986; Hussein, 2005).

#### **II.1. Prohibited Oviposition Efficiency by 6-BA:**

In the present study on *G. mellonella*, the oviposition efficiency of adult females was deleteriously prohibited, since the oviposition rate was drastically regressed, in a dosedependent course, after force-feeding of 3<sup>rd</sup> instar larvae on diets mixed with different concentrations of 6-BA. The present result was in corroboration with the reported results of prohibited oviposition of some insects by certain PGRs, such as the Asian citrus psyllid *Psylla citri* by PDC, Uniconazole or Mefluidide (Tsagkarakis *et al.*, 2012). Also, other studies showed that kinetin reduced the egg-laying capacity of the fruit flies (Kaur and Rup, 2002). Apart from PGRs, various plant-derived compounds had been reported to detrimentally inhibit the oviposition efficiency of some insect pests, such as *S. littoralis* by Nerolidol (Hamadah *et al.*, 2020) or Farnesol (Hamadah *et al.*, 2021); *S. oryzae* by Eugenol (Chaubey, 2016); the melon worm *Diaphania hyalinata* by Rotenone (Silva *et al.*, 2016) and the vinegar fly *Drosophila suzukii*by citral and eugenol (Eben *et al.*, 2020).

To interpret the inhibited oviposition efficiency of *G. mellonella* after force-feeding of the  $3^{rd}$  instar larvae on diet mixed with 6-BA, in the present study, it is important to point out that the reproduction in insects is mainly controlled by juvenile hormone (known as 'gonadotropic hormone' in adults). This hormone is responsible for the protein metabolism for egg maturation (Ghoneim *et al.*, 2014). On the other hand, ecdysteroids play a crucial role in the control of different processes involved in female reproduction, such as vitellogenesis and ovulation of matured eggs (Wigglesworth, 1984; Hagedorn, 1985). In the light of this information, the inhibited oviposition efficiency, in the current study, might be due to the interference of 6-BA with these hormones during the vitellogenesis *via* certain biochemical processes or might exert a reverse action to those exhibited by ecdysteroids which induce the neurosecretory cells to release a myotropic ovulation hormone (Smagghe *et al.*, 1996; Parween *et al.*, 2001). In addition, eggs might develop normally in ovaries of the adult females of *G. mellonella*, but they could not be lay, owing to the adversely deformed ovipositor by the action of 6-BA, or to the reduced mechanical strength (Moreno et al., 1994) or their reabsorption before oviposition (Zhou *et al.*, 2016).

# **II.2. Deteriorated Reproductive Capacity of** *G. mellonella* by 6-BA: **II.2.1. Prohibited Fecundity**:

After force-feeding of 3<sup>rd</sup> instar larvae of G. mellonella on diet mixed with 6-BA, the present study revealed a tremendous inhibitory effect of this PGR on the adult female fecundity (mean number of eggs/Q), in a dose-dependent manner. This result was in agreement with many reported results of inhibited fecundity of different insect species after treatment with various PGRs. For example, an early study was conducted by Robinson (1960) in which he recorded fecundity reduction of the pea aphid Acyrthosiphon pisum after feeding on broad bean treated withmaleic hydrazide (MH). Some years later, the 3<sup>rd</sup> instar nymphs of the same aphid species were fed on diets containing 0.5- 2.0% MH for three days leading to significantly reduced fecundity (Bhalla and Robinson, 1968). The fecundity of the black bean aphid Aphis fabae was inhibited when 1-15 ppm GA<sub>3</sub> was sprayed on Vicia fabae (Honeyborne, 1969). Chlormequat chloride (Chch) had been found to reduce the fecundity of several aphid species (Honeyborne, 1969; van Emden, 1969). After treatment of nymphs of the cotton stainer bug *Dysdercus cardinalis* with 2000 ppm chlormequat chloride (Chch), the emerged adult females had less fecundity (Carlisle et al., 1969). Feeding of S. littoralis larvae on a diet supplemented with  $GA_3$  (Salama and El-Sharaby, 1972) or Weedazol (3amino-1,2,4 triazol 25%) (Dimetry and Mansour, 1976) led to severe reduction of the adult fecundity. After addingGA<sub>3</sub> and ABA to a grass diet of the bigheaded grasshopper Aulocaraelliotti, significantly reduced fecundity was determined by Visscher (1980). The exogenous JA treatment of the wolfberry plant resulted in significantly reduced adult fecundity of the wolfberry aphid Aphis sp., in a dose-dependent course (Gong et al., 2010).

The lemon trees *Citrus volkameriana* were treated with PDC, Uniconazole, or Mefluidide. The rearing of Asian citrus psyllid *D. citri* on the treated trees led to significant reduction in adult fecundity (Tsagkarakis *et al.*, 2012). The injection of ABA into the haemocoel of *G. mellonella* resulted in a reduction of fecundity, at higher doses (Er and Keskin, 2016). Feeding of *G. mellonella* larvae on Mebendazole-treated diet from 1<sup>st</sup> instar larvae to the last instar resulted in significantly reduced fecundity of the producing female moths (Calıket al., 2016). Application of GA<sub>3</sub> (at 0.5 mg plant<sup>-1</sup>) on maize, as a host plant, significantly reduced the fecundity of *S. frugiperda* (Nagaratna *et al.*, 2021).

On the other hand, our result disagreed with few reported results of increasing fecundity of some insects after treatment with certain PGRs, such as *A. elliotti* after feeding on adiet treated with 10 and 20 mg/L doses of kinetin (Visscher, 1982); *D. melanogaster* after feeding on diet mixed with high doses ( $10^{-3}$  and  $10^{-4}$ M) of kinetin (Yeşilada and Bozcuk, 1996); the Silverleaf whitefly *Bemisiatabaci* after feeding on tomato seedlings sprayed with IAA (Di *et al.*, 2014) and the mulberry silkworm *Bombyx mori* after topical application of three doses of each GA<sub>3</sub> and Kinetin (Sepperumal and Sukumar, 2014).

To explicate the drastically inhibited adult fecundity of G. mellonella, after feeding of 3rd instar larvae on diet mixed with 6-BA, in the current study, this PGR might interfere with one or more physiological processes, from the ovarian follicle development to egg maturation. However, some suggestions could explain the presently prohibited fecundity, as follows. (1) 6-BA might cause some disorders in the ovaries, including cell death in the germarium, resorption of oocytes in the pre-vitellarium and vitellarium, formation of vitellin envelops and undue proliferation of follicle cells sometimes resulting in malformation of the whole ovary (Lucantoni et al., 2006; Khan et al., 2007). (2) The 6-BA might inhibit the development of some ovarioles and/or the synthesis and metabolism of proteinaceous constituents during the oogenesis (Salem et al., 1997). (3) The 6-BA might exert an inhibitory action on the ecdysone activity, the threshold of which is crucial for normal oogenesis (Terashima et al., 2005).(4) On the basis of hormonal regulation of insect reproduction, 6-BAmight disturb the production and/or function of the gonadotropic hormone (JH in adults) responsible for the synthesis of vitellogenins (volk precursors) and vitellogenesis (Di Ilioet al., 1999).(5) It may be acceptable to suggest that the prohibited fecundity may be due to theinhibitory effect of 6-BA on a synthesis of both DNA and RNA, suboptimal nutrition owing to reduced feeding, altered mating behaviour as a result of sublethal intoxication, or a combination of factors.

### II.2.2. Reduced Fertility:

Another informative parameter of the reproductive capacity is fertility (hatchingpercentage of laid eggs or egg viability) which was remarkably reduced after forcefeeding of 3<sup>rd</sup> instar larvae of G. mellonella on diet mixed with 6-BA, in the present study. Also, the sterility index increased parallel to the increasing concentration of 6-BA. This result was in accordance with many reported results of the fertility reduction in several insects after treatment with various PGRs. For example, feeding of the tobacco budworm (*Chloridea virescens*) larvae on a diet supplemented with GA<sub>3</sub> resulted in areduction of the hatching percentage of eggs laid by the successfully emerged adult females (Guerra, 1970). A similar result was recorded for S. littoralis by the same PGR (Salama and El-Sharaby, 1972). El-Ibrashy et al. (1976) injected Alar<sup>®</sup>85 into the newly moulted gregarious 4<sup>th</sup> instar nymphs of the desert locust Schistocerca gregaria and recorded complete sterility (zero fertility) in the treated locust. Adding GA<sub>3</sub> or ABA to a grass diet of A. elliotti significantly reduced the egg viability (Visscher, 1980). Treatment of the speckled rangeland grasshopper Arphia conspersa with a concentration of 60 mg/1 of ABA led to reduced fertility (Jurenka, 1982). Topical application of para-Aminobenzoic acid (vitamin  $B_x$ ) onto B. mori larvae caused a significant reduction in the egg hatchability (Pai et al., 1986). JA was reported to

reduce the egg hatchability in the brown planthopper *Nilapar vatalugens* (Senthil-Nathan *et al.*, 2009). Feeding of *G. mellonella* larvae on a mebendazole-treated diet from 1<sup>st</sup> instar larvae to the last instar resulted in significantly reduced hatchability of the laid eggs by adult females (Calık *et al.*, 2016). Application of GA<sub>3</sub> (at 0.5 mg plant<sup>-1</sup>) on maize, as a host plant, significantly reduced the fertility of *S. frugiperda* (Nagaratna et al., 2021). Recently, Çelik and Sak (2021) investigated the effects of kinetin against the smaller wax moth *Achroia grisella*. They found that kinetin, failed to affect egg fertility. On the contrary, our result disagreed with some exceptional cases of enhanced fertility of some insects by certain PGRs, such as the increasing fertility of the grasshopper *Aulocara elliotti* females after feeding on a diet mixed with 10 and 20 mg/L doses of kinetin (Visscher, 1982).

The detrimentally reduced fertility of G. mellonella adult females after forcefeeding of 3<sup>rd</sup> instar larvae on diet supplemented with 6-BA could be understood in the light of the following suggestions. (1) Maturation of the insect eggs depends basically on the vitellogenins, precursor materials of vitellins including proteins, lipids and carbohydrates, all of which are necessarily required for embryonic development (Soltani and Mazouni, 1992; Chapman, 1998). These materials are synthesized primarily by the fat body during the immature stages (Telfer, 2009) or by the ovary in situ (Indrasith et al., 1988). The 6-BAmight disturb the production of vitellogenins and/or accumulation in adult females, leading to the reduction of fertility. (2) The 6-BA might indirectly affect fertility via its disruptive effect on the opening of the intracellular spaces in follicular epithelium or generally inhibited the role of the gonadotropic hormone responsible for the regulation of vitellogenin deposition into oocytes (Davey and Gordon, 1996; Büyükgüzel, 2006). (3) The reduction in fertility might be due to the penetration of residual amounts of 6-BA in the adult females into their eggs and disturbance of embryonic cuticle synthesis. So, the fully mature embryos had weakened chitinous mouth parts that were insufficiently rigid to perforate the surrounding vitellin membrane and free from the eggs (Sallam, 1999; Sammour et al., 2008). (4) The reduced fertility might be due to the serious effect of 6-BA on survival of the developing embryos at certain stages, as recorded in decreasing hatching percentage.

#### II.2.3. Retarded Embryonic Development by 6-BA:

In insects, the incubation period of eggs can be used as an informative indicator of the embryonic developmental rate, i.e., a longer period usually denotes a slower rate of embryogenesis and vice versa. In the present study, 3rd instar larvae of G. mellonella were force-fed on diet mixed with 6-BA. The embryonic development in eggs laid by adult female moths was severely retarded with a slow rate since the incubation period of these eggs was significantly prolonged. To our knowledge, there was no reliable information in the available literature regarding the effects of PGRs on the embryonic development or incubation period of eggs, except a study by Uckan et al. (2011). Our result was in corroboration with their result of the prolonged incubation period of eggs laid by G. mellonella after rearing larvae on diet treated with >1.0 ppm of the GA<sub>3</sub>. On the other hand, the currently available literature contains very few studies examining the effect of some plant-derived compounds of the incubation period, such as the retarded embryonic development, or prolonged incubation period of eggs, after treatment of S. littoralis larvae with the Sesquiterpene compounds, Nerolidol (Hamadah et al., 2020) and Farnesol (Hamadah et al., 2021). The retarded embryonic development in G. mellonella, in the present study, might be due to an impairing effect of 6-BA on the ecdysteroids responsible for the regulation of certain stages of embryogenesis, especially those ecdysteroids originating from the ovary *in situ* (Chapman, 1998).

#### **Conclusion:**

Depending on the present results, the tested Cytokinin compound,6-Benzyladenine (6-BA) exhibited a considerable blocking effect on the adult emergence of *G. mellonella*, high insecticidal activity against adult moths, in addition to its prohibitory effect on oviposition efficiency, drastically reducing effect on fecundity and fertility, as well as it retarded the embryonic development in the laid eggs. Therefore, 6-BA may be an effective PGR being used in the IPM program against this serious pest of apiculture.

#### REFERENCES

- Abdel-Aal, A.E. (1996): Biological, histological and physiological effects of some insect growth regulators on the greasy cutworm, *Agrotis ipsilon* (Lepidoptera: Noctuidae).M.Sc. Thesis; Faculty of Science, Cairo University, Egypt.
- Abdellaoui K.; Ben Halima-Kamel M. and Ben Hamouda M.H. (2009): Physiological effects of gibberellic acid on the reproductive potential of *Locusta migratoria migratoria*. *Tunisian Journal of Plant Protection*, 4: 67-75.
- Abdellaoui, K.; Ben Halima-Kamel, M.; Acheuk, F.; Soltani, N.; Aribi, N. and Ben Hamouda, M.H. (2015): Effects of gibberellic acid on ovarian biochemical composition and ecdysteroid amounts in the migratory locust *Locusta migratoria* (Orthoptera, Acrididae). *International Journal of Pest Management*, 61: 68-72. https://doi.org/ 10.1080/09670874.2014.995746
- Acheuk, F.; Cusson, M. and Doumandji-Mitiche, B. (2012): Effects of a methanolic extract of the plant *Haplophyllum tuberculatum* and of teflubenzuron on female reproduction in the migratory locust, *Locusta migratoria* (Orthoptera: Oedipodinae). *Journal of Insect Physiology*, 58: 335-341.
- Al-Sharook, Z.; Balan, K.; Jiang, Y. and Rembold, H. (1991): Insect growth inhibitors from two tropical Meliaceae: Effects of crude seed extracts on mosquito larvae. *Journal* of Applied Entomology, 111: 425-430.
- Altuntaş, H. (2015): Determination of gibberellic acid (GA<sub>3</sub>)-induced oxidative stress in a model organism *Galleria mellonella* L. (Lepidoptera: Pyralidae). *Environmental Entomology (Physiology)*, 44(1): 100-105. DOI:10.1093/ee/nvu020.
- Altuntaş, H.; Kiliç, A.Y.; Uçkan, F. and Ergin, E. (2012): Effects of gibberellic acid on hemocytes of *Galleria mellonella* L. (Lepidoptera: Pyralidae). *Environmental Entomology*, 41(3): 688-696. https://doi.org/10.1603/EN11307
- Ambethgar, V. (2009): Potential of entomopathogenic fungi in insecticide resistance management (IRM): A review. *Journal of Biopesticides*, 2(2):177-193.
- Amer, M.S.; Ghoneim, K.S.; Al-Dali, A.G.; Bream, A.S.; Hamadah, Kh. Sh. (2004): Assessment of the activity of Margosan-0 and Jojoba against the house fly *Musca domestica* (Diptera: Muscidae). *Al-Azhar Bulletin of Science*, 15(2): 9-24.
- Ansari, M.S.; Ahmad, N. and Hasan, F. (2012): Potential of biopesticides in sustainable agriculture. In: "Environmental Protection Strategies for Sustainable Development" (Malik, A. and Grohmann, E., eds.). Dordrecht: Springer Science+Business Media BV: 529-595.
- Barbouche, N. and Ben Hamouda, M.H. (1986): Action des gibbérellines, hormones de croissancevégétales, sur la physiologie de la reproduction de *Ceratitis capitata* (Dipt. Tephritidae) In: Proceedings of II International Symposium of Fruit Flies/crete Greece, pp: 103-113.
- Bartlett, L. and Connor, E.F. (2014): Exogenous phytohormones and the induction of plant galls by insects. *Arthropod–Plant Interactions*, 8: 339–348.

- Bhalla, O.P. and Robinson, A.G. (1968): Effects of chemosterilants and growth regulators on the pea aphid fed on artificial diet. *Journal of Economic Entomology*, 68: 552-555.
- Bhatnagar, A. and Bareth, S.S. (2004): Development of low cost, high quality diet for greater wax moth, *Galleria mellonella* (Linnaeus). *Indian Journal of Entomology*, 66(3): 251-255.
- Bhatnagar, S.; Kumar, A. and Karnatak, A.K. (2012): Influence of synthetic plant growth stimulant, Miraculan on the survival and development of *Spodoptera litura* (Fab.)(Lepidoptera: Noctuidae). *Indian Forester*, 138(12): 1160-1163.
- Bhushan, S.; Gupta, S.; Sohal, S.K. and Arora, S. (2016): Assessment of insecticidal action of 3-Isothiocyanato-1-propene on the growth and development of *Spodoptera litura* (Fab.) (Lepidoptera: Noctuidae). *Journal of Entomology and Zoology Studies*, 4(5): 1068-1073.
- Body, M.; Kaiser, W.; Dubreuil, G.; Casas, J. and Giron, D. (2013): Leaf-miners co-opt microorganisms to enhance their nutritional environment. *Journal of Chemical Ecology*, 39: 969–977.
- Broughton, S.J.; Piper, M.D.; Ikeya, T.; Bass, T.M.; Jacobson, J.; Driege, Y.; Martinez, P.; Hafen, E.; Withers, D.J.; Leevers, S.J. and Partridge, L., (2005): Longer lifespan, altered metabolism, and stress resistance in *Drosophila* from ablation of cells making insulin-like ligands. *Proceedings of the National Academy of Sciences*, U.S.A, 102: 3105–3110.
- Bubán, T. (2020): The use of benzyladenine in orchard fruit growing: a mini review. *Plant Growrh Regulation*, 32:381–390.
- Büyükgüzel K. (2006): Malathion-induced oxidative stress in a parasitoid wasp: effect on adult emergence, longevity, fecundity, and oxidative and antioxidative response of *Pimpla turionellae* (Hymenoptera: Ichneumonidae). *Journal of Economic Entomology*, 99:1225-1234.
- Büyükgüzel, E. (2009): Evidence of oxidative and antioxidative responses by *Galleria mellonella* larvae to malathion. *Journal of Economic Entomology*, 102: 152-159.
- Büyükgüzel, E. and Kalender, Y. (2009): Exposure to streptomycin alters oxidative and antioxidative response in larval midgut tissues of *Galleria mellonella*. *Pesticide Biochemistry and Physiology*, 94: 112-118. https://doi.org/10.1016/ j.pestbp. 2009.04.008
- Calık, G.; Buyukguzel, K. and Buyukguzel, E. (2016): Reduced fitness in adults from larval, *Galleria mellonella* (Lepidoptera: Pyralidae) reared on media amended with the Antihelmintic, Mebendazole. *Journal of Economic Entomology*, 109(1): 182–187.
- Campbell, B.C.; Chan, B.G.; Creasy, L.L.; Dreyer, D.L.; Robin, L.B. and A. Wais, C. Jr. (1984): Bio-regulation of host plant resistance to insect. In "Bio-regulators; Chemistry and Action"(Ory, R.L. and Ritting, F.R., eds.). American Chemical Society, Symposium Series; pp.: 193-203.
- Carbone M.A.; Jordan K.W.; Lyman R.F.; Harbison S.T.; Leips J.; Morgan T.J.; DeLuca M.; Awadalla P. and Mackay T.F. (2006): Phenotypic variation and natural selection at catsup, a pleiotropic quantitative trait gene in *Drosophila*. *Current Biology*, 16: 912-919
- Carlisle, D.B.; Ellis, P.E.; Osborne, D.J. (1969): Effects of plant growth regulators on locusts and cotton stainer bugs. *Journal of the Science of Food and Agriculture*, 20: 391-393. https://doi.org/10.1002/jsfa. 2740200703
- Çelik, E. and Sak, O. (2021): Effects of kinetin on biological parameters and hemocytes of Achroiagrisella (Lepidoptera: Pyralidae). Archives of Biological Sciences, 72(2):181-192. https://doi.org/10.2298/ ABS200107012C

- Chamseddin, K.H.; Khan, S.; Nguyen, M.L.H. and Bauer, J. (2012): Takeout-dependent longevity is associated with altered juvenile hormone signaling. *Mechanisms of ageing and development*, 133: 11-12.
- Chandel, Y.S.; Sharma, S. and Verma, K.S. (2003): Comparative biology of the greater wax moth, *Galleria mellonella* L. and lesser wax moth, *Achoriagrisella*. *Forest Pest Management and Economic Zoology*, 11: 69-74.
- Chapman, R.F. (1998): The insects: structure and function. 4<sup>th</sup> ed. Cambridge: Cambridge University Press, pp: 116-118
- Chaubey, M.K. (2016): Insecticidal activities of *Cinnamomum tamala* (Lauraceae) essential oil against *Sitophilus oryzae* L. (Coleoptera: Curculionidae). *International Journal of Entomology Research*, 04 (03): 91-98.
- Chrominski, A.; S.N. Visscher, and R. Jurenka, (1982): Exposure to ethylene changes nymphal growth rate and female longevity in the grasshopper *Melanoplus* sanguinipes. Naturwissenschaften, 69:45.
- Davey, K.G, and Gordon, D.R.B. (1996): Fenoxycarb and thyroid hormones have JH-like effects on the follicle cells of *Locosta migratoria in vitro*. Archives of Insect Biochemistry and Physiology, 32: 613-626.
- Di, N.; Zhang, X.-Z.; Zhang, K.; Zhang, S.-Z. and Liu, T.-X. (2014): Influences of indole-3-acetic acid (IAA) spraying on *Solanum lycopersicum* plants and the infesting *Bemisia tabaci* (Gennadius) biotype B (Hemiptera: Aleyrodidae) whiteflies. *Acta Entomologica Sinica*, 57(7): 824-830.
- Di Ilio, V.; Cristofaro, M.; Marchini, D.; Nobili, P. and Dallai, R. (1999): Effects of a neem compound on the fecundity and longevity of *Ceratitis capitata* (Diptera: Tephritidae). *Journal of Economic Entomology*, 92:76-82.
- Dimetry, N.Z. and Mansour, M.H. (1976): Two synthetic plant growth regulators affecting the development and reproduction of the cotton leaf worm *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). *Journal of Applied Entomology*, 82(1-4): 435–440.
- Dreyer, D.L.; Campbell, B.C. and Jones, K.C. (1984): Effect of bioregulator-treated sorghum on greenbug fecundity and feeding behavior: Implication for host-plant resistance. Phytochemistry, 23: 1593-1596.
- Eben, A.; Sporer, F.; Vogt, H.; Wetterauer, P. and Wink, M. (2020): Search for alternative control strategies of *Drosophila suzukii* (Diptera: Drosophilidae): laboratory assays using volatile natural plant compounds. *Insects*, 2020, 11: 0811; doi:10.3390/ insects11110811
- El-Ibrashy, M.T.; Abdel-Hamid, M. and El-Refai, A. (1976): Ecdysones and plant growth regulators induce solitarious characters and reduce fertility in the desert locust. *Journal of Applied Entomology*, 19(3): 214–220.
- El-Wakeil, N. (2013): Botanical pesticides and their mode of action. *GessundePflanzen*, 65: 125-149.
- Er A. and Keskin M. (2016): Influence of abscisic acid on the biology and hemocytes of the model insect Galleria mellonella (Lepidoptera: Pyralidae). Annals of Entomological Society of America, 109: 244-251.https://doi. org/10.1093/ aesa/ sav122
- Er, A.; Taşkıran, D. and Sak, O. (2017): Azadirachtin-induced effects on various life history traits and cellular immune reactions of *Galleria mellonella* (Lepidoptera: Pyralidae). Archives of Biological Sciences, 69(2): 335-344. https://doi.org/10.2298/ABS160421108E.
- Farghaly, D.S.; El Sharkawy, A.Z.; Rizk, S.A. and Bader, N.F. (2017): Toxicological studies on the effect of Gamma radiation and some plant oils on greater wax moth *Galleria*

*mellonella* (Linnaeus) (Lepidoptera: Pyralidae). *Journal of Nuclear Technology in Applied Science*, 5: 143-152.

- Ghoneim, K. and Al-keridis, L.A. (2019): Effectiveness of Methoprene, a juvenile hormone analog, on adult performance and natality of the grey flesh fly *Parasarcophaga argyrostoma* (Diptera: Sarcophagidae). *African Entomology*, 27(1): 121–134.
- Ghoneim, K. and Bakr, R.F. (2018): Physiological activities of anti-Juvenile hormone agents against insects and their role for devising fourth generation insecticides: a comprehensive review. *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 11(3): 45-138.
- Ghoneim, K.S.; Mohamed, H.A. and Bream, A.S. (2000): Efficacy of the neem seed extract, Neemazal, on growth and development of the Egyptian cotton leafworrn, *Spodoptera littoralis* Boisd. (Lepidoptera: Noctuidae). *Journal of Egyptian German Societ of Zoology*, 33: 161-179.
- Ghoneim, K.; Tanani, M.; Hamadah, Kh.; Basiouny, A. and Waheeb, H. (2014): Inhibited reproductive capacity of Egyptian cotton leaf worm *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae) by the chitin synthesis inhibitor Novaluron. *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 7(2): 105-118.
- Ghoneim, K.; Hamadah, Kh.; Tanani, M.; Abdel-Khaliq, A. and Emam, D. (2019 a): Toxicity and disruptive impacts of the honeybee Apitoxin on growth and development of the greater wax moth, *Galleria mellonella* (Lepidoptera: Pyralidae). *Egyptian Academic Journal of Biological Sciences (F. Toxicology & Pest Control)*, 11(2): 97-106.
- Ghoneim, K.; Tanani, M.; Hamadah, Kh.; Abdel-Khaliq, A. and Emam, D. (2019 b): Deteriorated adult performance and reproduction of the greater wax moth, *Galleria mellonella* (Lepidoptera: Pyralidae) by the honey bee Apitoxin. Egyptian Academic *Journal of Biological Sciences (A. Entomology)*, 12(4): 95-108.
- Giron, D. and Glevarec, G. (2014): Cytokinin-induced phenotypes in plant-insect interactions: learning from the bacterial world. *Journal of Chemical Ecology*, 40: 826–835.
- Giron, D.; Frago, E.; Glevarec, G.; Pieterse, C.M. and Dicke, M. (2013): Cytokinins as key regulators in plant–microbe–insect interactions: connecting plant growth and defence. *Functional Ecology*, 27: 599–609.
- Glare, T.R.; Gwynn, R.L. and Moran-Diez, M.E. (2016): Development of biopesticides and future opportunities. In: "Microbial-Based Biopesticides: Methods and Protocols"(Glare, T.R. and Moran-Diez, M.E., eds.). New York: Springer Science+Business Media BV: 211-221.
- Gong, Y.Y.; Duan, L.Q.; Wang, A.Q.; Cui, R.J. and Qian, Y.S. (2010): Effects of exogenous jasmonic acid-induced resistance of wolfberry on the development and fecundity of the wolfberry aphid *Aphis* sp. *Acta Entomologica Sinica*, 53: 670–674.
- GraphPad InStat<sup>©</sup> v. 3.01 (1998): GraphPad Software, Inc.7825 Fay Avenue, Suite 230 La Jolla, CA 92037 USA. Available online at: http://www.graphpad.com/scientific-software/instat/
- Guerra, A.A. (1970): Effect of biologically active substances in the diet on development and reproduction of *Heliothis* spp. *Journal of Economic Entomology*, 63: 1518-1521.
- Gupta, G.; Yadav, S.R. and Bhattacharya, A.K. (2009): Influence of synthetic plant growth substances on the survivorship and developmental parameters of *Spilarctia obliqua* Walker (Lepidoptera: Arctiidae). *Journal of Pesticide Science*, 82: 41-46.
- Hagedorn, H.H. (1985): The role of ecdysteroids in reproduction. In: "Comprehensive Insect Physiology, Biochemistry and Pharmacology"(Kerkut, G.A., Gilbert, L.I., eds.). Pergamon, Oxford, vol. 8, pp: 205-262.

- Hamadah, Kh.; Ghoneim, K.; Waheeb, H. (2020): Impairing Effectiveness of Nerolidol, a Sesquiterpene Compound, on Adult Performance and Reproductive Potential of Egyptian Cotton Leafworm, Spodoptera littoralis (Lepidoptera: Noctuidae). Egyptian Academic Journal of Biological Sciences (A. Entomology), 13(2): 97-120.
- Hamadah, K.; Ghoneim, K.; Selim, Sh. and Waheeb, H. (2021): Bioactivity of Farnesol (a sesquiterpene compound) against the adult life parameters and reproductive potential of *Spodoptera littoralis* (Boisduval) (Lepidoptera: Noctuidae). *International Journal of Biosciences*, 18(3): 135-163. DOI: http://dx.doi.org/10.12692/ijb/18.3.135-163
- Herrera, J.M.; Zunino, M.P.; Dambolena, J.S.; Pizzolitto, R.P.; Ganan, N.A.; Lucini, E.I. and Zygadlo, J.A. (2015): Terpene ketones as natural insecticides against *Sitophilus zeamais*. *Industrial Crops and Products*, 70: 435-442.
- Honeyborne, C.H.B. (1969): Performance of Aphis fabae and Brevicorynebrassicae on plants treated with growth regulators. *Journal of the Science of Food and Agriculture*, 20: 388-390. https://doi.org/ 10.1002/jsfa.2740200702
- Hopkins, W.G. and Hüner, N.P. (2004): Introduction to Plant Physiology. 3<sup>rd</sup> Edition. John Wiley and Sons, Inc. 560pp.
- Hu, B.-Z.; Xu, Y.; Zheng, X.-R. and Shi, W.-P. (2012): Molt disruption and mortality of Locusta migratoria var. manilensis (Meyen) (Orthoptera: Acrididae) caused by insect growth regulators. African Journal of Biotechnology, 11(16): 3882-3887.
- Huang, S.-H.; Xian, J.-D.; Kong, S.-Z.; Li, Y.-C.; Xie, J.-H.; Lin, J.; Chen, J.-N.; Wang, H.-F. and Su, Z.-R. (2014): Insecticidal activity of pogostone against *Spodoptera litura* and *Spodoptera exigua* (Lepidoptera: Noctuidae). *Pest Management Science*, 70: 510–516.
- Hussein, K.T. (2005): Supressive effects of Calendula micrantha essential oil and gibberelic acid (PGR) on reproductive potential of the Mediterranean fruit fly *Ceratitis capitata* Wied. (Diptera: Tephritidae). Journal of Egyptian Society of Parasitology, 35: 365-377.
- Ilyas, A.; Khan, H.A.A. and Qadir, A. (2017): Effect of leaf extracts of some indigenous plants on settling and oviposition responses of peach fruit fly, *Bactrocerazonata* (Diptera: Tephritidae). *Pakistan Journal of Zoology*, 49: 1547-1553.
- Indrasith, L.; Sasaki, S.T.; Yaginuma, T.; Yamashita, O. (1988): The occurrence of premature form of egg-specific protein in vitellogenic follicles of *Bombyx mori*. *Journal of Comparative Physiology*, 158: 1-7.
- Jimenez-Peydro, R.; Gimeno-Martos, C.; Lopez-Ferror, J. Serrano- Delgado, C. and Moreno-Mari, J. (1995): Effects of the insect growth regulator, cyromazine, on the fecundity, fertility and offspring development of Mediterranean fruit fly, *Ceratitis capitata*Wied (Diptera, Tephritidae). *Journal of Applied Entomology*, 119: 435-438.
- Josephrajkumar, A.; Subrahmanyam, B. and Srinivasan, S. (1999): Plumbagin and azadirachtin deplete haemolymph ecdysteroid levels and alter the activity profiles of two lysosomal enzymes in the fat body of *Helicoverpa armigera* (Lepidoptera: Noctuidae). *European Journal of Entomology*, 96: 347-353.
- Jurenka, R.A. 1982): Studies on the embryonic development and embryonic diapause in Arphiaconspersa (Scudd.) and *Arphiapseu donietana* (Thomas) (Orthoptera, Acrididae) and the effects of plant growth hormones on reproduction and diapause. MSc. Thesis in Entomology, Montana State University, USA.
- Kartal, M.; Altun, M.L. and Kurucu, S. (2003): HPLC method for the analysis of harmol, harmalol, harmine and harmaline in the seeds of *Peganum harmala L. Journal of Pharmaceutical Biomedical Analysis*, 31: 263-269.

- Kaur, R. and Rup, P.J. (1999): Evaluation of gibberellic acid against immature stages of *Bactrocera cucurbitae* (Coquillett). *Journal of Insect Science*, 12: 9-14.
- Kaur, R. and Rup, P.J. (2002): Evaluation of regulatory influence of four plant growth regulators on the reproductive potential and longevity of melon fruit fly (*Bactroceracucurbitae*). *Phytoparasitica*, 30(3): 224-230. https://doi.org/ 10.1007/ BF03039991
- Kaur, R. and Rup, P.J. (2003): Influence of four plant growth regulators on development of the melon fruit fly, *Bactrocera cucurbitae* (Coquillett). *Insect Science and its Application*, 23(2):121-125.
- Kaur, N.; Gillett-Kaufman, J.L. and Buss, E.A. (2016): Effect of plant growth regulators on *Blissusin sularis* (Hemiptera: Blissidae). *Florida Entomologist*, 99(3):557-558. https://doi.org/10.1653/024.099.0336
- Kefford, B.J.; Zalizniak, L.; Warne, M.S.J. and Nugegoda, D. (2008): Is the integration of hormesis and essentiality into ecotoxicology now opening Pandora's box? *Environmental Pollution*, 151:516-23.
- Khan, M.; Hossain, M.A. and Islam, M.S. (2007): Effects of neem leaf dust and a commercial formulation of a neem compound on the longevity, fecundity and ovarian development of the melon fly, *Bactocera cucurbitae* (Coquillett) and the oriental fruit fly, *Bactrocera dorsalis* (Hendel) (Diptera: Tephritidae). *PakistanJournal of Biological Sciences*, 10: 3656-3661.
- Kumar, S. and Gupta, K.K. (2017): Influence of Farnesol on growth and development of *Dysdercus koenigii*. 19<sup>th</sup> International Conference of Entomology, 2017, held at Paris, France, October, 19-20, 2017.
- Kunbhar, S.; Rajput, L.B.; Ahmed, G.A.; Akber, C.G. and Sahito, J.G.M. (2018): Impact of botanical pesticides against sucking insect pests and their insect predators in brinjal crop. *Journal of Entomology and Zoology Studies*, 6: 83–87.
- Kwadha, C.A.; Ongamo, G.O.; Ndegwa, P.N.; Raina, S.K. and Fombong, A.T. (2017): The biology and control of the greater wax moth, *Galleria mellonella*. *Insects*, 9, 8(2): 61. doi: 10.3390/insects8020061
- Liao, M.; Xiao, J.J.; Zhou, L.J.; Yao, X.; Tang, F.; Hua, R.M.; Wu, X.W. and Cao, H.Q. (2017): Chemical composition, insecticidal and biochemical effects of *Melaleuca alternifolia* essential oil on the *Helicoverpa armigera*. *Journal of Applied Entomology*, 141: 721–728. https://doi.org/10.1111/jen.12397
- Lingampally, V.; Solanki, V.R.; Kaur, A. and Raja, S.S. (2013): Andrographolide- an effective insect growth regulator of plant origin against *Tribolium confusum* (Duval). *International Journal of Current Research*, 5(1): 22-26.
- Łozowicka, B.; Jankowska, M. and Kaczyński, P. (2012): Pesticide residues in *Brassica* vegetables and exposure assessment of consumers. *Food Control*, 25(2): 561–575.
- Lucantoni, L.; Giusti, F.; Cristofaro, M.; Pasqualini, L.; Esposito, F.; Lupetti, P. and Habluetzel, A. (2006): Effects of a neem extract on blood feeding, oviposition and oocyte ultrastructure in *Anopheles stephensi* Liston (Diptera: Culicidae). *Tissue Cell*, 38: 361-371.
- Maazoun, A.M.; Ben Hlel, T.; Hamdi, S.H.; Belhadj, F.; Ben Jemâa, J.M. and Marzouki, M.N. (2017): Screening for insecticidal potential and acetylcholinesterase activity inhibition of Urginea maritima bulbs extract for the control of Sitophilus oryzae (L.). Journal of Asia-Pacific Entomology, 20: 752-760.
- Mahdi, M.N. and Mohammed, A.J. (2017): Detection of oxamyl and imidacloprid pesticide residues in some Iraqi vegetables. *IOSR Journal of Agriculture and Veterinary Sciences*, 10(7): 67-80.

- Majid, G.J.; Ali, S.; Seyed, A.M.; Iraj, A. and Foad, M. (2011): Effects of the exogenous application of auxin and cytokinin on carbohydrate accumulation in grains of rice under salt stress. *Plant Growth Regulators*, 65: 305–313.
- Mangena, P. (2020): Role of Benzyladenine seed priming on growth and physiological and biochemical response of soybean plants grown under high salinity stress condition. *International Journal of Agronomy*, 5: 1-5. DOI: 10.1155/ 2020/ 8847098
- Marouf, A.E. (2020): Efficacy of nano chitosan and mandarin crust oil on some biological aspects of cotton leafworm, *Spodoptera littoralis* (Boisd.). *International Journal of Entomology Research*, 5(1): 89-95.
- Mendonça, J.M.A.; Carvalho, G.A.; Guimarães, R.J.; Reis, P.R. and Rocha, L.C.D. (2006): Produtosnaturais e sintéticos no controlede *Leucoptera coffeella* (Guérin-Mèneville&Perrottet, 1842) (Lepidoptera:Lyonetiidae) e seusefeitos sobre a predação por vespas. *Ciência e Agrotecnologia*, 30: 892-899.
- Metwally, H.M.S.; Hafez, G.A.; Hussein, M.A.; Salem, H.A.; Saleh, M.M.E. (2012): Lowcost artificial diet for rearing the greater wax moth, *Galleria mellonella* L. (Lepidoptera: Pyralidae) as a host for entomopathogenic nematodes. *Egyptian Journal of Biological Pest Control*, 22(1): 15-18
- Moreno, D.S.; Martinez, A.J. and Riviello, M. S. (1994): Cyromazine effects on the reproduction of *Anastreph aludens* (Diptera: Tephritidae) in the laboratory and in the field. *Journal of Economic Entomology*, 87: 202–211.
- Moroney, M.J. (1956): Facts from figures (3<sup>rd</sup> ed.). Penguin Books Ltd., Harmondsworth. Middle Sex.
- Müller, A.M. and Theron, K.I. (2018): The use of 6-benzyladenine to increase same-season branching on one-year-old lateral extension growth of third-leaf apple trees. *Acta Horticulturae*, 1206: 63-68 DOI: 10.17660/ActaHortic.2018.1206.9
- Nagaratna, W.; Kalleshwaraswamy, C.M.; Dhananjaya, B.C.; Sharanabasappa, N. and Prakash, N.B. (2021): Effect of silicon and plant growth regulators on the biology and fitness of fall armyworm, *Spodoptera frugiperda*, a recently invaded pest of maize in India. Silicon, 11 pp. https://doi.org/10.1007/s12633-020-00901-8
- Nandi, P.S. and Chakravarty, K. (2011): Juvenoids and anti-Juvenoids as third generation pesticide to control lepidopteran field crop pests. *Indian Streams Research Journal*, 1(6): 15pp.
- Nitin, K.; Kumar, K.D.; Kumar, M.V. and Sanjay, P. (2012): Effect of economical modification in artificial diet of greater wax moth *Galleria mellonella* (Lepidoptera: Pyralidae). *Indian Journal of Entomology*, 74(4): 369-374.
- Nurunnaher, A.; Rafiqul, I.M.; Abdul, K.M. and Tofazzal, H. (2014): Alleviation of drought stress in maize by exogenous application of gibberellic acid and cytokinin. *Journal of Crop Science and Biotechnology*, 17:41–48.
- Osman, E.E.; Rarwash, I. and El- Samadisi, M.M. (1984): Effect of the anti-moulting agent "Dimilin" on the blood picture and cuticle formation in *Spodoptera littoralis* (Boisd.) larval. *Bulletin of Entomological Society of Egypt (Economic Series)*, 14: 3-46.
- Owain, E.R.; Franzmann, B.; Thackray, D. and Micic, S. (2008): Insecticide resistance and implications for future aphid management in Australian grains and pastures: A review. *Australian Journal of Experimental Agriculture*, 48:1523
- Pai, I.K.; Hegde, S.N. and Krishnamurthy, N.B. (1986): Effect of paraminobenzoic acid on the hatchability in silkworm, *Bombyx mori* L. *The Indian Zoologist*, 10: 7-13.
- Parween, S.; Faruki, S.I. and Begum, M. (2001): Impairment of reproduction in the red flour beetle, *Tribolium castaneum* (Herbst.) (Coleoptera: Tenebrionidae) due to larval feeding on triflumuron-treated diet. *Journal of Applied Entomology*, 125: 1-4.

- Prado, S.G. and Frank, S.D. (2013): Tritrophic effects of plant growth regulators in an aphidparasitoid system. *Biological Control*, 66: 72-76. https://doi.org/ 10.1016/ j.biocontrol.2013.03.019
- Rehman, J.U.; Jilani, G.; Khan, M.A. and Kanvil, S. (2009): Repellent and oviposition deterrent effects of indigenous plant extracts on peach fruit fly, *Bactrocer azonata* Saundera (Diptera: Tephritidae). *Pakistan Journal of Zoology*, 49: 101-108.
- Robinson, A.G. (1960): Effect of maleic hydrazide and other plant growth regulators on the pea aphid, *Acyrthosiphon pisum* (Harris), caged on broad bean, *Vicia faba* L. *The Canadian Entomologist*, 92(7): 494-499. https://doi.org/10.4039/Ent92494-7
- Roch, M.; Varela, M.C.; Taglialegna, A. and Rosato, A.E. (2020): Tedizolid is a promising antimicrobial option for the treatment of *Staphylococcus aureusinfections* in cystic fibrosis patients. *Journal of Antimicrobial Chemistry*, 75:126–134. DOI: 10.1093/jac/dkz418
- Rortais, A.; Arnold, G.; Dorne, J.L.; More, S.J.; Sperandio, G.; Streissl, F. and Verdonck, F. (2017): Risk assessment of pesticides and other stressors in bees: principles, data gaps and perspectives from the European Food Safety Authority. *Science of the Total Environment*, 587: 524-537.
- Rup, P.J.; Kaur, R. and Kaur, J. (1998): Effect of gibberellic acid (GA<sub>3</sub>) on the protein, lipid and carbohydrate contents of banana fruit fly, *Zaprionus paravittiger* larvae. *Insect Science and its Application*, 18: 145-148. https://doi.org/10. 1017/ S1742758400007785
- Rup, P.J.; Sohal, S.K.; Sohi, R.; Kaur, G.; Sandhu, N.; Gurm, S.K.; Dhingra, P. and Wadhwa, S.K. (2000): Influence of PGRs on carbohydrate content in *Lipaphis erysimi* (Kalt.). *Indian Journal of Experimental Biology*, 38: 1066-1068.
- Rup, P.J.; Sohal, S.K.; Kaur, G. and Sharma, R. (2002): The possible role of five enzymes in the metabolism of kinetin in mustard aphid, *Lipaphis erysimi* (Kaltenbach). *Journal of Aphidology*, 16:1-8.
- Saad, M.M.G.; Abou-Taleb, H.K. and Abdelgaleil, S.A.M. (2018): Insecticidal activity of monoterpenes and phenylpropenes against *Sitophilus oryzae* L. and their acetylcholinesterase and adenosine triphosphatases inhibitory effects. *Applied Entomology and Zoology*, 53: 173–181.
- Said, S.M.; Hammam, M.A. and Abd-El Kader, S.K. (2019): Insecticidal activity against the greater wax moth (*Galleria mellonella* L.) and chemical composition of five plant essential oils. *Menoufia Journal of Plant Protection*, 4: 145 161.
- Salama, H.S. and El-Sharaby, A.M. (1972): Gibberellic acid and β-sitosterol as sterilants of the cotton leafworm, *Spodoptera littoralis*Boisduval (Lepidoptera: Noctuidae). *Experientia*, 28: 413-414.
- Salem, H.; Smagghe, G. and Degheele, D. (1997): Effects of tebufenozide on oocyte growth in *Plodia interpunctella*. Medical. Faculty. Landbouww. *Gent University*, 62(1): 9-13.
- Salisbury, F.B. and Ross, C.W. (1992): Plant Physiology. Belmont, CA: Wadsworth. pp. 357-407, 531-548.
- Sallam, M.H. (1999): Effect of Diflubenzuron on embryonic development of the acridid, *Heteracris littoralis. Journal of Egyptian Geram Society of Zoology*, 30(E): 17-26.
- Sammour, E.A; Kandit, M.A. and Abdel-Aziz, N.F. (2008): The reproductive potential and fat of clorfluazuron and lufenuron against cotton leafworm, *Spodoptera littoralis* (Boisd). *American-Eurasian Journal of Agriculture and Environmental Science*, 4(1): 62-67.

- Senthil-Nathan, S.; Kalaivani, K.; Choi, M.Y. and Paik, C.H. (2009): Effects of jasmonic acid-induced resistance in rice on the plant brownhopper, *Nilapar vatalugens* Stal (Homoptera: Delphacidae). *Pesticide Biochemistry and Physiology*, 95: 77-84
- Sepperumal, U. and Sukumar, S. (2014): Impact of kinetin and gibberellic acid on the commercial characteristics of the silkworm, *Bombyx mori* L. (Lepidoptera: Bombycidae). *Journal of Microbiology and Biotechnology Research*, 4(1):11-20.
- Shiwani, B. and Karnatak, A. K. (2012): Effect of gibberellic acid on the developmental profile of *Spodoptera litura* (Fab.). Pantnagar Journal of Research, 10(1): 18-21.
- Silva, R.D.A.; Santos, J.L.; Oliveira, L.S.; Soares, M.R.S.; Santos, S.M. S.D. (2016): Biostimulants on mineral nutrition and fiber quality of cotton crop. *Revista Brasileira de Engenharia Agrícola e Ambiental*, 20 (12), 1062-1066. http://dx.doi.org/10.1590/1807-1929/agriambi. v20n12p1062-1066.
- Singh, H. and Bhattacharya A.K. (2001): Role of plant growth regulators on the developmental profile of *Spodoptera litura*: Effect of plant growth stimulants. *Indian Journal of Entomology*, 63(3): 329-339.
- Smagghe, G. and Degheele, D. (1992): Effects of the non-steroidal ecdysteroid agonist, RH-5849 on reproduction of *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). *Phytoparasitica*, 48: 23-29.
- Smagghe, G.; Salem, H.; Tirry, L. and Degheele, D. (1996): Action of a novel insect growth regulator tebufenozide against different developmental stages of four stored product insects. *Parasitica*, 52: 61-69.
- Soltani, N. (1984): Effects of ingested diflubenzuronon the longevity and peritrophic membrane of adult mealworms (*Tenebrio molitor* L.). *Pesticide Science*, 15: 221-225.
- Soltani, N. and Mazouni, A. (1992): Diflubenzuron and oogenesis in the codling moth, *Cydiapomonella. Pesticide Science*, 34: 257-261.
- Sosa, A.; Diaz, M.; Salvatore, A.; Bardon, A.; Borkosky, S. and Vera, N. (2019): Insecticidal effects of Vernonanthur anebularum against two economically important pest insects. Saudi Journal of Biological Sciences, 26: 881–889.
- Sparks, T.C. and Nauen, R. (2015): IRAC: mode of action, classification and insecticide resistance management. *Pesticide Biochemistry and Physiology*, 121, 122–128.
- Stamm, P.; Ramamoorthy, R. and Kumar, P.P. (2011): Feeding the extra billions: strategies to improve crops and enhance future food security. *Plant Biotechnology Reports*, 5: 107-120.
- Tanani, M. and Ghoneim, K. (2018): Disruptive effects of certain chitin synthesis inhibitors on adult life parameters and reproductive potential of the pink bollworm, *Pectinophora gossypiella* (Saunders)(Lepidoptera: Gelechidae). *Egyptian Academic Journal of Biological Sciences (A. Entomology)*, 11(5): 79-102.
- Telfer, W.H. (2009): Egg formation in Lepidoptera. Journal of Insect Science, 9: 50. (insectscience.org/9.50).
- Terashima, J.; Takaki, K.; Sakurai, S. and Bownes, M. (2005): Nutritional status affects 20hydroxyecdysone concentration and progression of oogenesis in *Drosophila melanogaster*. Journal of Endocrinology, 187: 69-79.
- Toppozada, A.; Abd-allah, S. and El-Defrawi, M.E. (1966): Chemosterilization of larvae and adults of the Egyptian cotton leafworm, *Prodenialitura* by apholate, metepa and hempa. *Journal of Economic Entomology*, 59: 1125-1128.
- Trigo, J.R.; Campos, S. and Pereira, A.M. (1988): Presença de alcalóidespirrolizidinicosem Ageratum conyzoides L. In: "Simposio de PlantasMedicinais do Brasil", Sao Paulo. (Resumos). p. 13.

- Tsagkarakis, A.E.; Rogers, M.E. and Spann, T.M. (2012): Applications of plant growth regulators to container-grown citrus trees affect the biology and behavior of the Asian citrus psyllid. *Journal of the American Society for Horticultural Science*, 137: 3-10.
- Uçkan, F.; Haftacı, İ. and Ergin, E. (2011): Effects of indol-3-acetic acid on biological parameters of the larval endoparasitoid *Apantales galleriae* (Hymenoptera: Braconidae). *Annals of the Entomological Society of America*, 104: 77-82.
- Uçkan, F.; Soydabaş, H.K. and Özbek, R. (2014): Effect of Indol-3-acetic acid on the biochemical parameters of *Achoria grisella* hemolymph and *Apanteles galleriae* larva. *Pakistan Journal of Biotechnology*, 11(2): 163-171.
- van Emden, H.F. (1969): Plant resistance to *Myzus persicae* induced by a plant regulator and measured by aphid relative growth rate. *Entomologia Experimentalis et Applicata*, 12:125-131.
- Visscher, N.S. (1980): Regulation of grasshopper fecundity, longevity and egg viability by plant growth hormones. *Experientia*, 36:130–131.
- Visscher, S.N. (1982): Plant growth hormones affect grasshopper growth and reproduction. In: Proceedings of the 5<sup>th</sup> International Symposium of Insect-Plant Relationships (Visser, J.H. and Minks, A.K., editors). Wageningen: Pudoc, Pp: 57-62.
- Wigglesworth, V.B. (1984): Insect Physiology. 8th ed., Chapman & Hall, London, 191 pp.
- Wilson, T.G. (2004): The molecular site of action of juvenile hormone and juvenile hormone insecticides during metamorphosis: how these compounds kill insects. *Journal of Insect Physiology*, 50: 111–121.
- Yarahmadi, F.; Moassadegh, M.; Soleymannejadian, E.; Saber, M. and Shishehbor, P. (2009): Assessment of acute toxicity of abamectin, spinosad and chlorpyrifos to *Thrips tabaci* Lindeman (Thysanoptera: Thripidae) on sweet pepper by using two bioassay techniques. *Asian Journal of Biological Sciences*, 2(3):81-87.
- Yeşilada, E. and Bozcuk, A.N. (1996): The effects of ABA and kinetin on the developmental period of *Drosophila melanogaster*. *Turkish Journal of Biology*, 20:29-35.
- Yeşilada, E.; Bozcuk, A.N.; Bozcuk, S. and Topcuoğlu, F. (1996): The effects of abscisic acid (ABA) and kinetin on sex-ratio and adult morphology of *D. melanogaster*. *Turkish Journal of Biology*, 20:171-178.
- Zhang, Z.; Yang, T.; Zhang, Y.; Wang, L. and Xie, Y. (2016): Fumigant toxicity of monoterpenes against fruitfly, *Drosophila melanogaster*. *Industrial Crop Products*, 81: 147–151.